

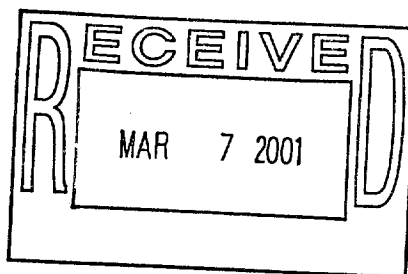
Supplementary Material: Experimental Procedures, Characterization Data and ^1H NMR spectra

**Rapid Entry into the Cryptophycin Core via an Acyl- β -Lactam Macrolactonization:
Total Synthesis of Cryptophycin-24**

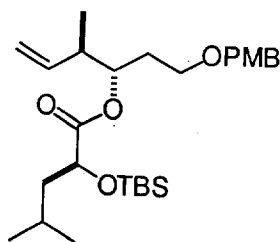
MariJean Eggen, Sajiv, K. Nair, Gunda I. Georg*

Department of Medicinal Chemistry

University of Kansas, Lawrence, KS 66045



(1*S*,2*R*)-1-(2-(4-Methoxybenyloxy)ethyl)-2-methyl-3-butenyl (2*S*)-4-methyl-2-[(*tert*-butyldimethylsilyl)oxy]pentanoate (10).



To (*tert*-Butyldimethylsilyl)oxy (2*S*)-4-methyl-2-[(*tert*-butyldimethylsilyl)oxy]pentanoate (6.5 g, 18 mmol) in 30 mL CH₂Cl₂ was added 15 drops of DMF. The solution was cooled to 0 °C and oxalyl chloride (3.2 mL, 36 mmol) was added dropwise using an addition funnel. Vigorous bubbling was observed which gradually subsided. After addition was complete, the mixture was warmed to rt and stirred at rt for 4.5 h. The mixture was concentrated *in vacuo* and used in the next step.

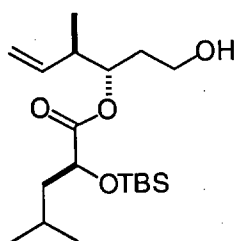
The crude acid chloride 6 was added dropwise to a solution of DMAP (2.2 g, 18 mmol) in CH₂Cl₂ (10 mL) at 0 °C. In a second flask, the alcohol (1.5 g, 6 mmol) was dissolved in CH₂Cl₂ (5 mL) and TEA (1.7 mL, 12 mmol) was added. The alcohol solution was added dropwise to the first. The reaction was stirred at 0 °C for 45 min at which time TLC indicated disappearance of alcohol 7. The reaction was quenched with saturated aqueous NaHCO₃ and Et₂O. The aqueous layer was extracted with Et₂O. The combined organics were dried (MgSO₄) and concentrated. Flash chromatography (hexanes to 95:5 hexanes:EtOAc) provided ester 10 as an oil (2.22 g, 78%): [α]_D -42° (c 1.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.25-7.23 (d, *J* = 8 Hz, 2H), 6.87-6.85 (d, *J* = 8 Hz, 2H), 5.77-5.68 (m, 1H), 5.08-5.05 (m, 2H), 5.02-5.01 (d, 5 Hz, 1 H), 4.43-4.40 (d, *J* = 11.4 Hz, 1H), 4.37-4.34 (d, *J* = 11.4 Hz, 1H), 4.20-4.17 (dd, *J* = 9, 4 Hz, 1H), 3.79 (s, 3H), 3.48-3.36 (m, 2H), 2.43-2.38 (m, 1H), 1.85-1.83 (d, *J* = 7 Hz, 1H), 1.82-1.80 (d, *J* = 7Hz, 1H), 1.75 (m buried, 1H), 1.64-1.57 (ddd, *J* = 5, 9, 14 Hz, 1H), 1.50-1.44 (ddd, *J* = 4, 9, 14 Hz, 1H), 1.00-0.99 (d, *J* = 7 Hz, 3H), 0.924-0.907 (d, *J* = 7 Hz, 3H), 0.91

(buried, 3H), 0.90 (s, 9H), 0.07 (s, 3H), 0.04 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 173.8, 159.1, 139.0, 130.4, 129.2 (2C), 115.8, 113.7 (2 C), 74.3, 72.7, 70.7, 66.5, 55.2, 44.3, 41.6, 31.6, 25.7 (3C), 24.1, 23.4, 21.7, 18.2, 15.7, -4.6, -5.3; IR (film) 3050, 2920, 2820, 1730, 1625, 1590, 1570 cm^{-1} ; MS (EI) m/z 477 (M-1, 25), 421 (15), 362 (50), 201 (50).

Three step synthesis of 11 from 10.

1. DDQ-deprotection of 10 to form intermediate alcohol 14.

(1S)-3-Hydroxy-1-[(1R)-1-methyl-2-propenyl]propyl] (2S)-4-methyl-2-
[(*tert*-butyldimethylsilyl)oxy]pentanoate (14).

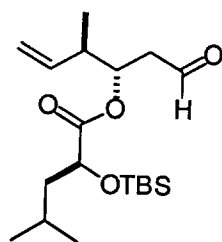


Ester 10 (276 mg, 0.577 mmol) was dissolved in CH_2Cl_2 (2 mL) and H_2O (0.1 mL). The mixture was cooled to 0 °C and DDQ (144 mg, 0.635 mmol) was added. After 30 min at 0 °C, the bath was removed and the mixture was stirred an additional 90 min at rt. Additional CH_2Cl_2 was added and the organics were washed with saturated aqueous NaHCO_3 solution. The combined organics were dried (MgSO_4), filtered and concentrated. Flash chromatography (hexanes to 85:15 hexanes:EtOAc) provided the desired alcohol 14 (176 mg, 85%) as an oil (often a mixture of alcohol and aldehyde by-product were utilized in the next reaction): ^1H NMR (400 MHz, CDCl_3) δ 5.76-5.68 (ddd, J = 8, 10, 17.8 Hz, 1H), 5.08-4.99 (m, 3H), 4.25-4.22 (dd, J = 4, 9 Hz, 1H), 3.65-3.60 (m, 1H), 3.51-3.45 (ddd, J = 4, 10, 11.5 Hz, 1H), 2.42-2.37 (m, 2H), 1.84-1.75 (m, 2H), 1.71-1.58 (m, 2H), 1.54-1.47 (ddd, J = 4, 9, 13.5 Hz, 1H), 1.03-1.01 (d, J = 7 Hz, 1H), 0.924-0.91 (d, J = 7 Hz, 3H), 0.916-0.90 (d, J = 7 Hz, 3H), 0.89 (s, 9H), 0.07 (s, 3H), 0.04 (s, 3H); ^{13}C

NMR (100 MHz, CDCl₃) δ 175.2, 139.0, 116.1, 74.5, 70.7, 58.5, 44.4, 41.9, 34.7, 25.7 (3 C), 24.2, 23.4, 21.7, 18.2, 16.2, -4.7, -5.3; IR (film) 3420, 3060, 2920, 2890, 2820, 1730 cm⁻¹.

2. Oxidation of DDQ deprotected alcohol **14** to intermediate aldehyde **15**.

(1S)-1-(1-Methyl-2-propenyl)-3-oxopropyl (2S)-4-methyl-2-[(*tert*-butyldimethylsilyl)oxy]pentanoate (**15**).

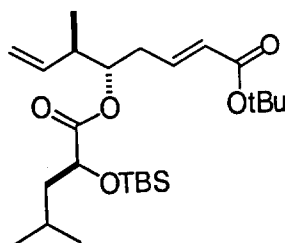


The alcohol **14** (95 mg, 0.265 mmol) was dissolved in CH₂Cl₂ (2.7 mL) and Dess-Martin periodinane (169 mg, 0.397 mmol) was added. After 2 h, a 1:1 mixture of saturated aqueous Na₂S₂O₃ and NH₄Cl solutions was added. After 10 min of vigorous stirring, the mixture was diluted with CH₂Cl₂ and H₂O. The aqueous phase was extracted with CH₂Cl₂. The combined extracts were washed with saturated aqueous NaHCO₃ solution, dried (MgSO₄), filtered and concentrated. Product **15** was a colorless oil (76 mg, 81%): ¹H NMR (400 MHz, CDCl₃) δ 9.7 (dd, *J* = 2.5, 1.4 Hz, 1H), 5.76-5.67 (ddd, *J* = 17, 10.4, 8 Hz, 1H), 5.39-5.34 (dt, *J* = 8, 5 Hz, 1H), 5.11-5.09 (m, 1H), 5.10-5.06 (m, 1H), 4.19-4.16 (dd, *J* = 4, 9 Hz, 1H), 2.68-2.62 (ddd, *J* = 17, 8, 2.5 Hz, 1H), 2.62-2.56 (ddd, *J* = 17, 5, 1.4 Hz, 1H), 2.53-2.48 (m, 1H), 1.80-1.74 (m, 1H), 1.63-1.56 (ddd, *J* = 14, 9, 5 Hz, 1H), 1.48-1.42 (ddd, *J* = 14, 9, 4 Hz, 1H), 1.04-1.03 (d, *J* = 7 Hz, 3H), 0.92-0.89 (buried, 6H), 0.89 (s, 9H), 0.07 (s, 3H), 0.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 199.2, 173.7, 138.2, 116.9, 71.7, 70.7, 45.5, 44.2, 41.3, 25.7 (3 C), 24.1, 23.4, 21.7, 18.2, 15.5, -4.7, -5.4; IR (film) 3750, 2940, 2910, 2840, 1740, 1715 cm⁻¹;

MS (EI) m/z 357 ($M + 1$, 100), 316, 303, 201, 189; HRMS (FAB) $C_{19}H_{37}O_4Si$ ($M+H$)
calc'd for: 357.2461, found 357.2463.

3. Wittig reaction of intermediate aldehyde 15 to form 11.

tert-Butyl (5*S*,6*R*,2*E*)-6-methyl-5-(4-methyl-(2*S*)-2-[(*tert*-butyldimethylsilyl)oxy] pentanoyloxy)octa-2,7-dienoate (11).

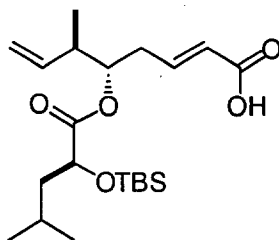


The aldehyde 15 (80 mg, 0.22 mmol) was dissolved in CH_2Cl_2 (2.2 mL) and cooled to 0 °C. (*tert*-Butoxycarbonylmethylene) triphenylphosphorane (101 mg, 0.27 mmol) was added. After 30 min at 0 °C and 90 min at rt, the reaction was concentrated and purified by flash chromatography (95:5 hexane:EtOAc) to provide 11 as a colorless oil (96 mg, 94 %): $[\alpha]_D^{25}$ (c 0.50, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$) δ 6.77-6.69 (dt, $J = 7$, 15.5 Hz, 1H), 5.79-5.75 (d, $J = 15.5$ Hz, 1H), 5.75-5.66 (m, 1H), 5.08 (br s, 1H), 5.07-5.03 (m, 1H), 4.99-4.95 (app q, $J = 6$ Hz, 1H), 4.19-4.16 (dd, $J = 4$, 9 Hz, 1H), 2.45-2.38 (m, 3H), 1.81-1.74 (m, 1H), 1.63-1.57 (ddd, $J = 5$, 9, 13.5 Hz, 1H), 1.48-1.42 (buried, 1H), 1.45 (s, 9H), 1.02-1.00 (d, $J = 7$ Hz, 3H), 0.92-0.89 (buried, 6H), 0.89 (s, 9H), 0.06 (s, 3H), 0.02 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 173.8, 165.3, 142.3, 138.6, 125.9, 116.3, 80.2, 75.3, 70.7, 44.3, 41.2, 34.1, 28.1 (3 C), 25.7 (3 C), 24.1, 23.4, 21.7, 18.2, 16.0, -4.6, -5.4; IR (film) 3070, 2935, 2910, 2840, 1740, 1700, 1645 cm^{-1} ; MS (EI) m/z 399 (9), 341 (9), 267 (20), 201 (100), 189 (100); HRMS (FAB) calc'd for $C_{25}H_{47}O_5Si$ ($M+H$): 455.3193, found 455.3171.

Two step synthesis of 12 from 11.

1. Deprotection of ester **11** to intermediate acid **16**.

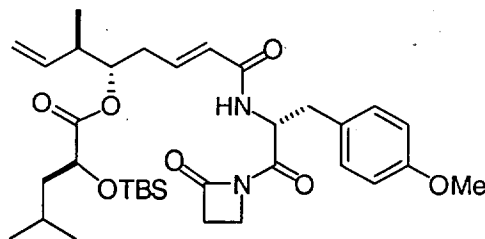
(5S,6R,2E)-6-Methyl-5-((2S)-4-methyl-2-[(*tert*-butyldimethylsilyl)oxy]pentanoyloxy)octa-2,7-dienoic acid (16**).**



The *tert*-butyl ester **11** (40 mg, 0.088 mmol) was dissolved in 0.1 mL of CH₂Cl₂ and cooled to 0 °C. A TFA solution (1 M in CH₂Cl₂, 1.1 mL) was added via syringe. The ice bath was removed after 2 h and the reaction was continued with vigorous stirring. After 6 h, the reaction was found to be complete by TLC. Toluene (0.10 mL) was added and the reaction was concentrated under vacuum to provide acid **16** as a colorless oil (29 mg, 83%): ¹H NMR (400 MHz, CDCl₃) δ 6.99-6.92 (dt, *J* = 8, 15.5 Hz, 1H), 5.88-5.84 (br d, *J* = 15.5 Hz, 1H), 5.74-5.65 (ddd, *J* = 8, 10.5, 17 Hz, 1H), 5.09-4.99 (m, 3H), 4.19-4.16 (dd, *J* = 4, 9.5 Hz, 1H), 2.49-2.38 (m, 3H), 1.79-1.74 (m, 1H), 1.63-1.56 (ddd, *J* = 4.5, 9.5, 13.5 Hz, 1H), 1.45-1.38 (ddd, *J* = 4, 9.5, 13.5 Hz, 1H), 1.02-1.00 (d, *J* = 7 Hz, 3H), 0.91-0.90 (d, *J* = 4.5 Hz, 3H), 0.89-0.88 (d, *J* = 4.5 Hz, 3H), 16.52 (s, 9H), 6.05 (s, 3H), 5.76 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.8, 146.7, 138.5, 123.3, 116.5, 75.0, 70.7, 44.4, 41.3, 34.5, 25.7 (3 C), 24.1, 23.4, 21.7, 18.2, 16.1, -4.7, -5.4; IR (film) 3060, 3000, 2930, 2900, 2840, 1740, 1715, 1680, 1640 cm⁻¹; MS (EI) *m/z* 399 (M+H, 3), 341 (3), 267 (4), 247 (10), 231 (30); HRMS (FAB) calc'd for (M+H) C₂₁H₃₉O₅Si: 399.2576, found 399.2559.

2. Amide formation of **12** from acid **16** and salt **9**.

(1S,3E)-4-(*N*-((1*R*)-1-((4-Methoxybenzyl)-2-oxo-2-(2-oxoazetidiny)ethyl)carbamoyl)-1-((1*R*)-1-methyl-2-propenyl)but-3-enyl 4-methyl-2-[(*tert*-butyldimethylsilyl)oxy]pentanoate (12**).**



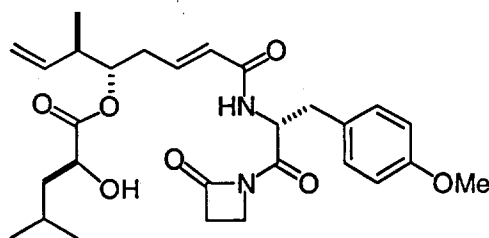
The *N*-Boc-4-methoxy (D)-phenylalanine (120 mg, 0.41 mmol) was dissolved in CH₃CN (4 mL) and cooled to 0 °C. 2-Azetidinone (50 mg, 0.69 mmol), DIEA (142 μ L, 0.81 mmol) and HBTU (262 mg, 0.69 mmol) were added consecutively. After 20 min, the ice bath was removed and the mixture was stirred 1 h. The reaction was quenched with saturated aqueous NH₄Cl solution and extracted with EtOAc. The combined organics were dried (MgSO₄), filtered and concentrated. Flash chromatography (80:20 to 50:50 hexanes:EtOAc) provided (2*R*)-(tert-butoxy)-*N*-((4-methoxyphenyl)methyl)-2-oxo-2-(2-oxoazetidinyl)ethylformamide as a cream colored solid (85 mg, 60%): [α]_D -56° (c 1.0, CHCl₃); m.p. 107-109°; ¹H NMR (400 MHz, CDCl₃) δ 7.12-7.10 (d, *J* = 8.6 Hz, 2H), 6.82-6.80 (d, *J* = 8.6 Hz, 2H), 5.05 (br s, 2H), 3.76 (s, 3H), 3.64-3.59 (m, 1H), 3.51-3.46 (m, 1H), 3.08-2.98 (m, 3H), 2.82-2.78 (m, 1H), 1.35 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 164.7, 159.4, 155.7, 130.8 (2C), 128.1, 114.3 (2C), 80.4, 56.2, 55.7, 37.5, 36.9, 36.4, 28.7 (3C); IR (film) 3390, 1790, 1705, 1690 cm⁻¹; MS(FAB) *m/z* 349 (M+H, 8), 293 (84), 249(25), 231 (76), 121 (100); HRMS (FAB, NBA) calc'd for C₁₈H₂₅N₂O₅ (M+H): 349.1763, found 349.1782.

This compound (150 mg, 0.431 mmol) was dissolved in 4N HCl in dioxane (4 mL) and stirred for 1.5 h. A precipitate formed and the reaction was concentrated to provide a light tan salt **9**.

Acid **16** (40 mg, 0.10 mmol) and salt **9** (40 mg, 0.140 mmol) were placed in a flask and CH₃CN (1 mL) was added. The solution was cooled to 0 °C and DIEA (52 mL, 0.30 mmol) was added. When solution was clear, HBTU (57 mg, 0.15 mmol) was added and the reaction stirred for 30 min at 0 °C and rt for 90 min. Saturated NaHCO₃ solution and

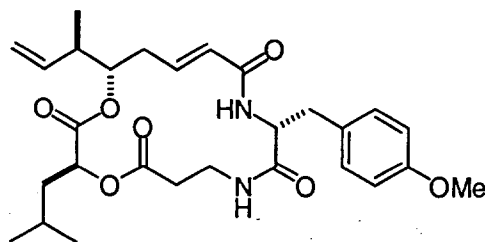
CH₂Cl₂ were added. The aqueous layer was further extracted with CH₂Cl₂. The combined organics were dried (MgSO₄), filtered and concentrated. Flash chromatography (70:30 to 40:60 hexane/EtOAc) provided the product **12** as a thick oil (40 mg, 64%): ¹H NMR (400 MHz, CDCl₃) δ 7.10-7.07 (d, *J* = 8.6 Hz, 2H), 6.82-6.80 (d, *J* = 8.6 Hz, 2H), 6.75-6.68 (dt, *J* = 7.5, 15 Hz, 1H), 5.98-5.96 (br d, *J* = 7.5 Hz, 1H), 5.82-5.78 (d, *J* = 15 Hz, 1H), 5.73-5.65 (m, 1H), 5.39-5.34 (app q, *J* = 7 Hz, 1H), 3.06-5.01 (m, 2H), 4.94-4.89 (app q, *J* = 6 Hz, 1H), 4.18-4.15 (dd, *J* = 4, 9 Hz, 1H), 3.77 (s, 3H), 3.64-3.59 (m, 1H), 3.53-3.48 (m, 1H), 3.15-3.10 (dd, *J* = 6, 14 Hz, 1H), 3.08-2.99 (m, 2H), 2.97-2.91 (dd, *J* = 7.5, 14 Hz, 1H), 2.43-2.39 (m, *J* = 7 Hz, 3H), 1.80-1.73 (m, 1H), 1.62-1.55 (ddd, *J* = 5, 9, 13.5 Hz, 1H), 1.47-1.41 (ddd, *J* = 4, 9, 13.5 Hz, 1H), 0.99-0.98 (d, *J* = 7 Hz, 3H), 0.91-0.89 (d, *J* = 7 Hz, 3H), 0.90-0.88 (d, *J* = 7 Hz, 3H), 0.88 (s, 9H), 0.04 (s, 3H), 0.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.8, 169.5, 164.7, 164.2, 158.8, 140.1, 138.6, 130.3 (2 C), 127.2, 125.6, 116.3, 114.0 (2 C), 75.4, 70.7, 55.2, 54.5, 44.3, 40.8, 36.7, 36.6, 36.0, 34.0, 25.7 (3 C), 24.1, 23.4, 21.7, 18.2, 16.3, -4.7, -5.4; IR (film) 3280, 3060, 2920, 2900, 2825, 1770, 1730, 1690, 1660, 1620 cm⁻¹; MS (EI) 629 (M⁺, 7), 572 (5), 457 (5), 383 (20), 284 (20), 231 (45), 121 (75); HRMS (FAB) calc'd for C₃₄H₅₂N₂O₇Si 629.3622, found 629.3622.

(1*S*,3*E*)-4-(*N*-((1*R*)-1-((4-Methoxybenzyl)-2-oxo-2-(2-oxoazetidiny)ethyl)carbamoyl)-1-((1*R*)-1-methyl-2-propenyl)but-3-enyl (2*S*)-2-hydroxy-4-methylpentanoate (5).



The silyl ether **12** (6.0 mg, 9.5 μ mol) was dissolved in CHCl_3 (1 mL, dried over activated 4 Å molecular sieves). $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (10 μ L, 76.3 μ mol) was added via syringe. After 1 h, CHCl_3 (5 mL) and saturated, aqueous NaHCO_3 (2 mL) were added. The organics were dried (MgSO_4), filtered and concentrated to provide clean **5** as an oil (4.4 mg, 88%): ^1H NMR (400 MHz, CDCl_3) δ 7.10-7.08 (d, J = 8.6, 2H), 6.83-6.81 (d, J = 8.6, 2H), 6.71-6.63 (dt, J = 7.5, 15 Hz, 1H), 6.07-6.05 (br d, J = 7.5, 1H), 5.81-5.77 (d, J = 15, 1H), 5.71-5.62 (ddd, J = 8, 10.4, 17 Hz, 1H), 5.36-5.33 (app q, J = 6 Hz, 1H), 5.08-5.02 (m, 2H), 4.97-4.93 (app q, J = 6 Hz, 1H), 4.14-4.12 (br s, 1H), 3.77 (s, 3H), 3.64-3.61 (m, 1H), 3.54-3.49 (m, 1H), 3.16-3.11 (dd, J = 6, 14 Hz, 1H), 3.08-3.03 (m, 2H), 2.95-2.90 (dd, J = 8, 14 Hz, 1H), 2.45-2.40 (m, 3H), 1.91-1.84 (m, 1H), 1.52-1.48 (m, 2H), 1.00-0.99 (d, J = 6.9 Hz, 3H), 0.95-0.93 (d, J = 7 Hz, 3H), 0.94-0.92 (d, J = 7 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.4, 169.6, 164.9, 164.3, 158.8, 139.7, 138.4, 130.3 (2C), 127.2, 125.8, 116.6, 114.0 (2C), 76.4, 69.0, 55.2, 54.6, 43.5, 41.1, 36.6 (2C), 36.0, 34.1, 24.5, 23.3, 21.4, 16.2.

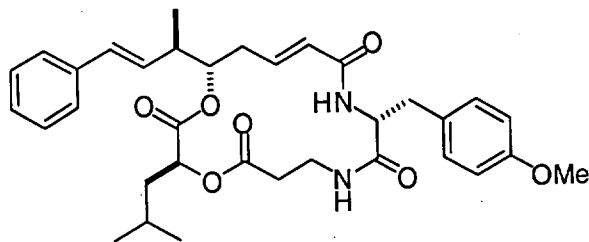
Dephenyldesepoxyarenastatin A (**4**).



Alcohol **5** (4.4 mg, 8.6 μ mol) was dissolved in CH_2Cl_2 (2 mL). Bu_4NCN (21 mg, 77.0 μ mol) was dissolved in CH_2Cl_2 (2 mL) and both were stirred with flame activated crushed 4 Å molecular sieves for 1 h. The Bu_4NCN solution was transferred via syringe dropwise to the solution of **5**. After 16 h, the reaction was filtered through celite and concentrated. Column chromatography (50:50 to 75:25 EtOAc:hexanes) provided clean product **4** (3.0 mg, 68%): ^1H NMR (400 MHz, CDCl_3) δ 7.11-7.09 (d, J = 8.6 Hz, 2H), 7.05-7.02 (t, J

= 5.7 Hz, 1H), 6.81-6.79 (d, J = 8.6 Hz, 2H), 6.71-6.63 (ddd, J = 5, 10.3, 15 Hz, 1H), 5.84-5.82 (d, J = 8 Hz, 1H), 5.75-5.71 (d, J = 14 Hz, 1H), 5.69-5.65 (m, 1H), 5.08 (br s, 1H), 5.05-5.04 (d, J = 6 Hz, 1H), 5.02-4.97 (ddd, J = 2, 5, 11 Hz, 1H), 4.94-4.91 (dd, J = 4, 9.5 Hz, 1H), 4.72-4.66 (m, 1H), 3.76 (s, 3H), 3.54-3.47 (m, 1H), 3.47-3.40 (m, 1H), 3.16-3.11 (dd, J = 6, 14.4 Hz, 1H), 3.03-2.97 (dd, J = 7.6, 14.4 Hz, 1H), 2.44-2.28 (m, 2H), 1.76-1.66 (m, 1H), 1.48-1.40 (m, 1H), 1.04-1.02 (d, J = 7 Hz, 3H), 0.92-0.90 (d, J = 6.4 Hz, 3H), 0.88-0.87 (d, J = 6.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.8, 170.82, 170.76, 165.7, 158.5, 141.8, 138.6, 130.2 (2C), 128.5, 124.9, 116.5, 114.1 (2C), 76.8, 71.4, 55.2, 54.3, 42.4, 39.8, 36.1, 35.2, 34.2, 32.5, 24.5, 22.9, 21.5, 16.6; IR (film) 3420, 3300, 2990, 1740, 1675, 1525 cm^{-1} ; MS (FAB) 515.3 (M+H); HRMS (FAB) calcd for $\text{C}_{28}\text{H}_{39}\text{O}_7\text{N}_2$ (M+H) 515.2757, found 515.2775.

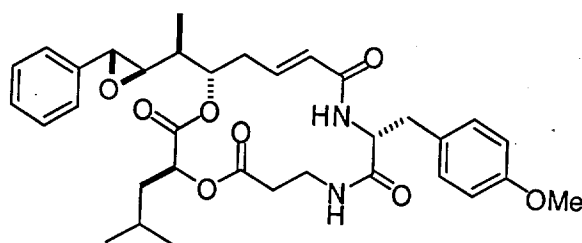
Desepoxyarenastatin A (13).



Olefin 4 (17 mg, 33 μmol) was dissolved in CH_3CN (0.33 mL) in a dry sealed tube. The solution was flushed with argon and iodobenzene (4.0 mL, 36 μmol), $\text{Pd}(\text{OAc})_2$ (1.1 mg, 5.0 μmol) and TEA (46 mL, 330 μmol) were added. The tube was sealed and placed in a 80-85 $^\circ\text{C}$ oil bath with vigorous stirring overnight. After 20 h, the solution was filtered and purified by silica gel chromatography to obtain product **13** as a solid (3 mg, 31% based on recovered starting material) and unreacted starting material **4** (6 mg): $[\alpha]_{\text{D}}^{20} +27^\circ$ (c 0.80, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.33-7.15 (m, 5H), 7.11-7.09 (d, J = 8.6 Hz, 2H), 7.05-7.02 (t, J = 5.5 Hz, 1H), 6.81-6.78 (d, J = 8.6 Hz, 2H), 6.73-6.66 (ddd, J = 4.7, 10.5, 15 Hz, 1H), 6.41-6.37 (d, J = 15.8 Hz, 1H), 6.03-5.96 (dd, J = 8.8, 15.8 Hz, 1H), 5.77-5.75 (d, J = 7.9 Hz, 1H), 5.75-5.71 (d, J = 15 Hz, 1H), 5.06-5.01 (ddd, J =

2, 6.6, 11 Hz, 1H), 4.91-4.88 (dd, $J = 3.6, 10$ Hz, 1H), 4.73-4.67 (m, 1H), 3.76 (s, 3H), 3.54-3.48 (m, 1H), 3.46-3.39 (m, 1H), 3.15-3.11 (dd, $J = 6, 14.4$, 1H), 3.04-2.98 (dd, $J = 7.5, 14.4$ Hz, 1H), 2.57-2.52 (m, 3H), 2.39-2.30 (m, 1H), 1.78-1.57 (m, 3H), 1.35-1.27 (m, 1H), 1.13-1.11 (d, $J = 6.8$ Hz, 3H), 0.73-0.72 (d, $J = 6.4$ Hz, 3H), 0.70-0.69 (d, $J = 6.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.8, 170.9, 170.8, 165.6, 158.5, 141.7, 136.7, 131.8, 130.2 (2C), 128.6 (2C), 128.5, 127.5, 126.1 (2C), 125.0, 114.1 (2C), 76.6, 71.5, 55.2, 54.3, 42.2, 39.7, 36.4, 35.2, 34.2, 32.4, 24.3, 22.6, 21.2, 17.2; IR (film) 3365, 3260, 2940, 1725, 1710, 1665 cm^{-1} ; MS (FAB) m/z 591.3 (M+H); HRMS (FAB) calcd for $\text{C}_{34}\text{H}_{43}\text{N}_2\text{O}_4$ (M+H) 591.3070, found 591.3069.

Arenastatin A (1).



Olefin 13 (5.0 mg, 8.5 μmol) was reacted as previously reported¹ with dimethyldioxirane² to obtain a 2:1 mixture (*de* was determined by HPLC Phenomenex Hypersil 5 μ , C18, 150 $\times$ 3.2 mm, 254 nm, 3:2 $\text{CH}_3\text{CN}:\text{H}_2\text{O}$, 0.5 mL/min, retention time: $\beta = 7.2$ min, $\alpha = 7.9$ min) of epoxide diastereomers (3.9 mg, 76%): $[\alpha]_{\text{D}}^{20} +37^\circ$ (c 0.10, CHCl_3).¹

References:

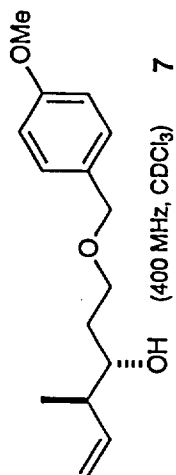
- (1) White, J. D.; Hong, J.; Robarge, L. A. *J. Org. Chem.* **1999**, *64*, 6206-6216.
- (2) Adam, W.; Bialas, J.; Hadjirapoglou, L. *Chem. Ber.* **1991**, *124*, 2377.

Current Data Parameters
NAME HJE
EXPNO 2
PROCNO 1

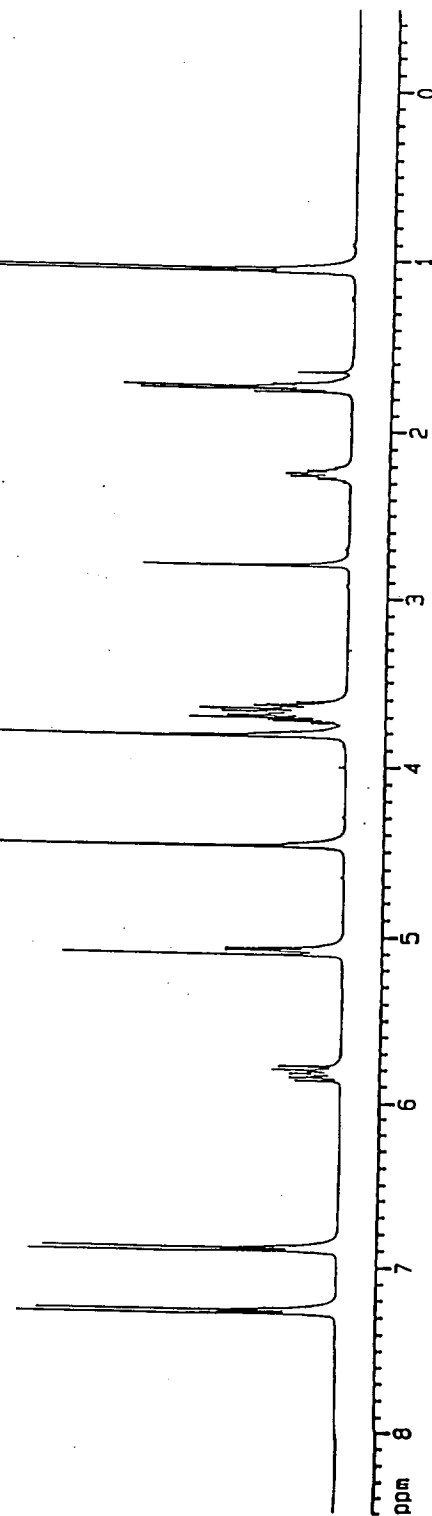
F2 - Acquisition Parameters
Date_ 990121
Time 11.07
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zg30
TO 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 181
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec
P1 7.70 usec
DE 4.50 usec
SF01 400.1320007 MHz
NUC1 1H
PL1 -6.00 dB

F2 - Processing parameters
SI 16384
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
F1P 8.500 ppm
F1 3401.10 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCH 0.45000 ppm/cm
HZCH 180.05850 Hz/cm



S11

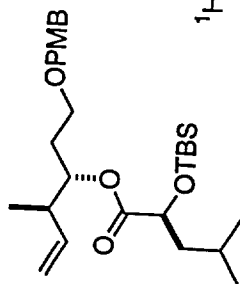


Current Data Parameters
NAME SN-0-7719-13
EXPNO 1
PROCNO 1

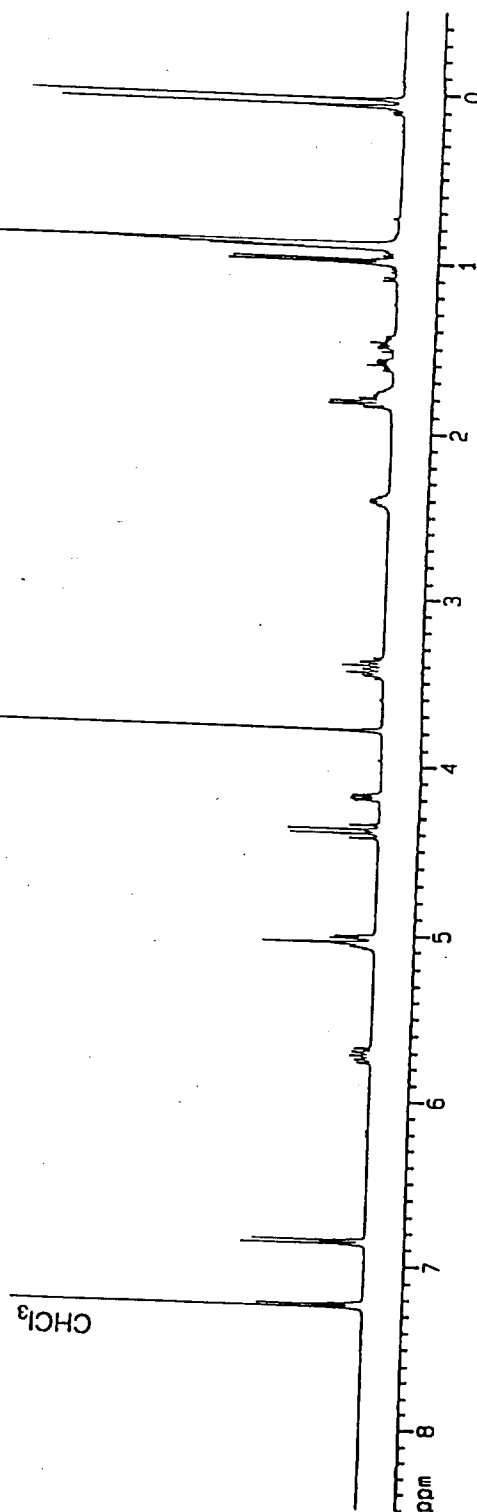
F2 - Acquisition Parameters
Date_ 990204
Time 14.06
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zg30
TO 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 456.1
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec
P1 7.70 usec
DE 4.50 usec
SF01 400.132007 MHz
NUC1 1H
PL1 -6.00 dB

F2 - Processing parameters
SI 16384
SF 400.1300135 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

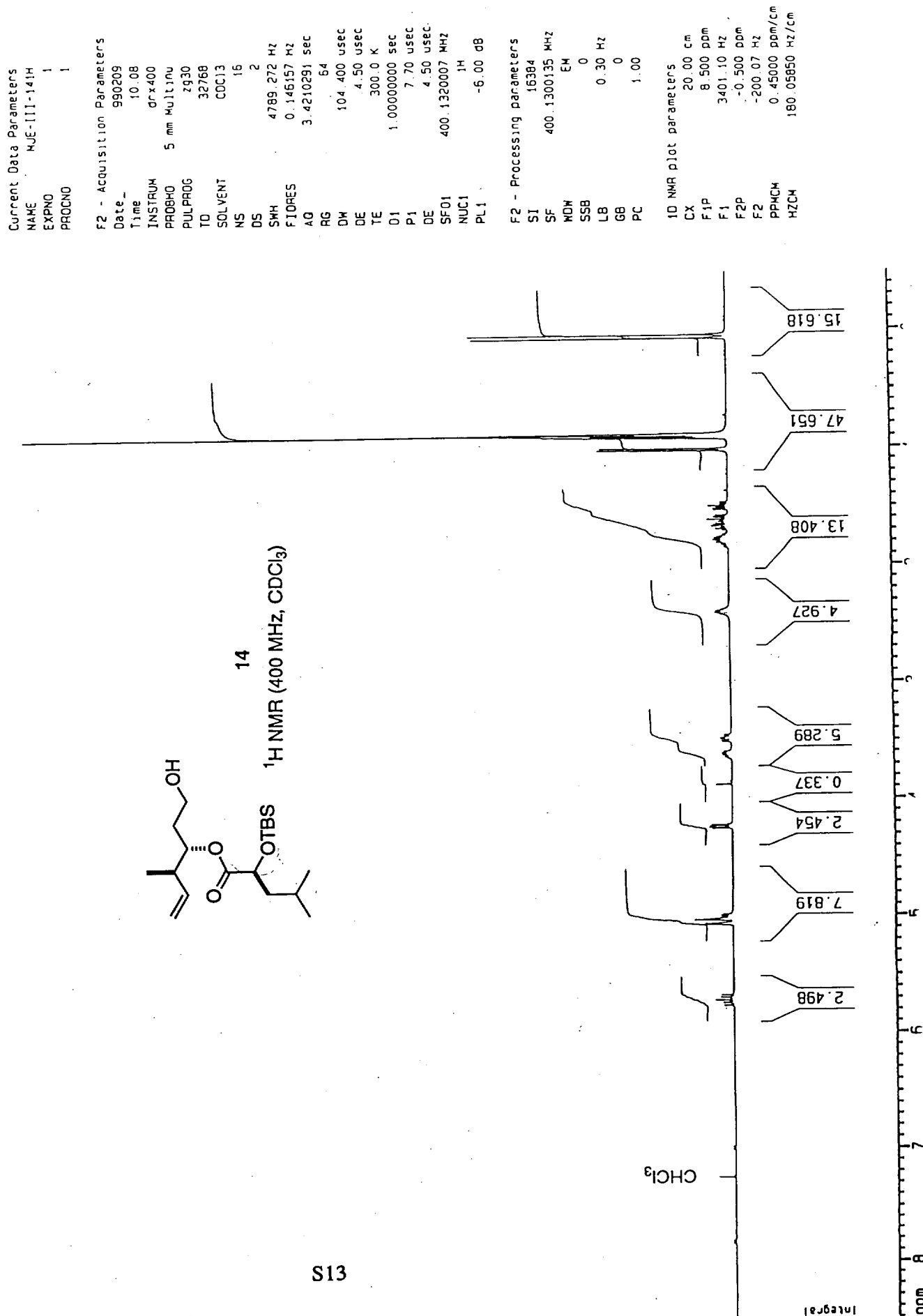
10 NMR plot parameters
CX 20.00 cm
F1P 8 500 ppm
F1 3401.10 Hz
F2P -0 500 ppm
F2 -200.07 Hz
PP4CH 0 45000 ppm/cm
HZCH 180.05850 Hz/cm



10
¹H NMR (400 MHz, CDCl₃)



S12



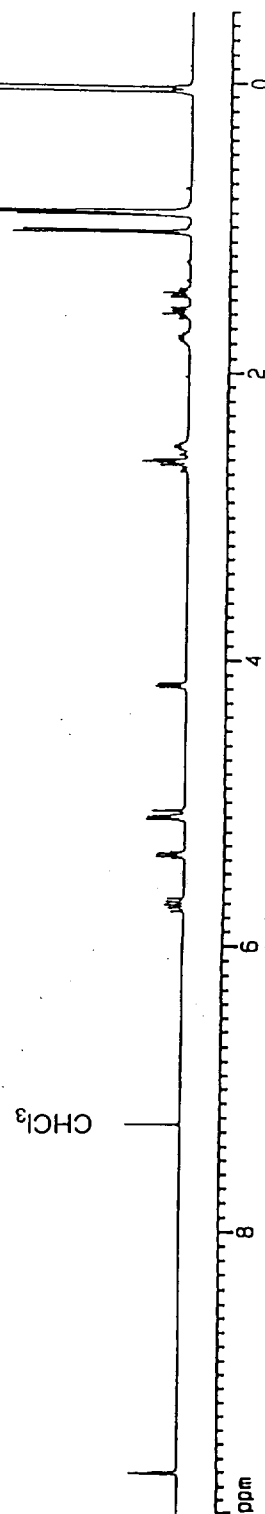
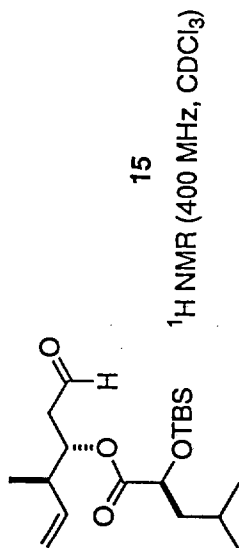
Current Data Parameters
 NAME MJE-III-142H
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 990209
 Time 17.06
 INSTRUM drx400
 PROBHD 5 mm MUltiPnu
 PULPROG zg30
 TO 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SMH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 161.3
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 P1 7.70 usec
 DE 4.50 usec
 SF01 400.1320007 MHz
 NUC1 1H
 PL1 -6.00 dB

F2 - Processing parameters
 SI 16384
 SF 400.1300135 MHz
 HSW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PP4CH 0.52500 ppm/cm
 HZCH 210.06825 Hz/cm



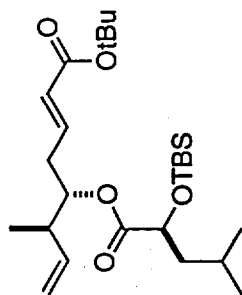
S14

Current Data Parameters
NAME MJE-111-143
EXPNO 1
PROCNO 1

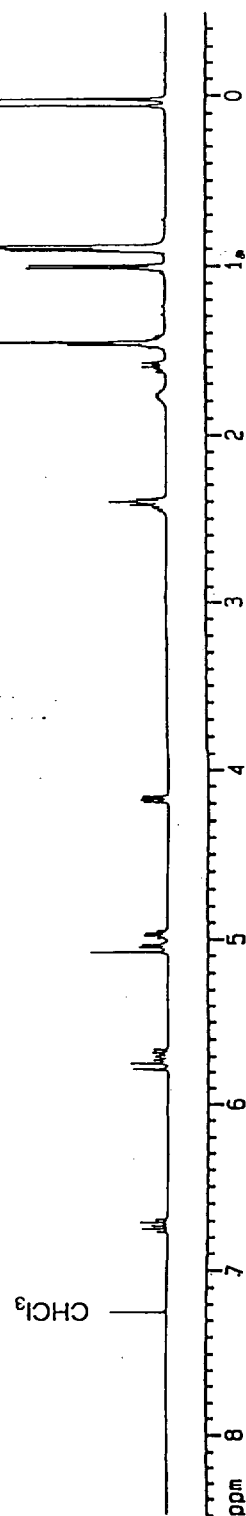
F2 - Acquisition Parameters
Date_ 990212
Time 16.53
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 128
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec
P1 7.70 usec
DE 4.50 usec
SFO1 400.1320007 MHz
NUC1 1H
PL1 -6.00 dB

F2 - Processing parameters
SI 16384
SF 400.1300135 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
F1P 8.500 ppm
F1 3401.10 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCH 0.45000 ppm/cm
HZCH 180.05850 Hz/cm



11

¹H NMR (400 MHz, CDCl₃)

S15

Current Data Parameters
 NAME HJE-III-148H
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 990305
 Time 13.15

INSTRUM grx400
 PROBD 5 mm HJ111nu
 PULPROG zg30

TD 32768
 SOLVENT CDCl3
 NS 16

DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz

AQ 3.4210291 sec
 RG 512
 DR 104.400 usec

DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec

P1 7.70 usec
 DE 4.50 usec
 SF01 400.1320007 MHz

NUC1 1H
 PL1 -6.00 dB

F2 - Processing Parameters
 SI 16384
 SF 400.1300000 MHz

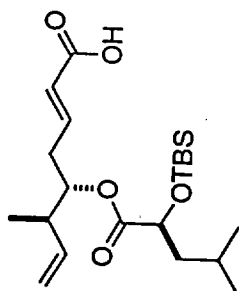
WDW EM
 SSB 0
 LB 0.30 Hz

GB 0
 PC 1.00

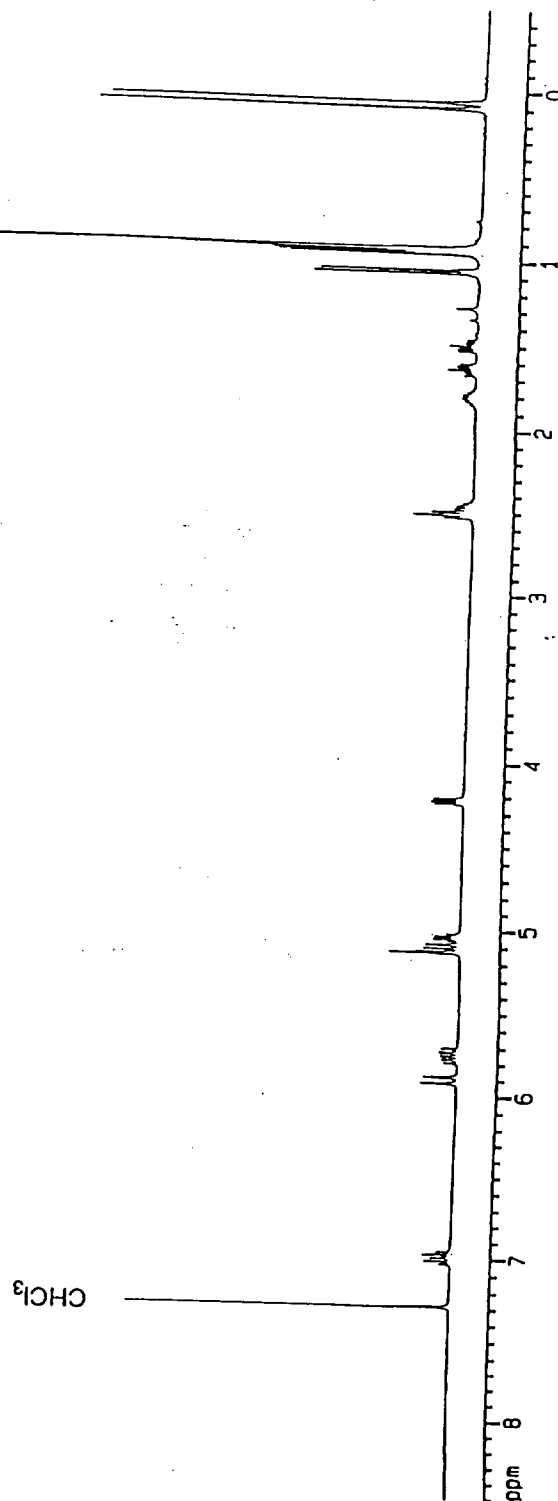
1D NMR plot parameters
 CX 20.00 cm
 F1P 8.500 ppm

F1 3401.10 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz

PP4CH 0.45000 ppm/cm
 HZCH 180.05850 Hz/cm



16

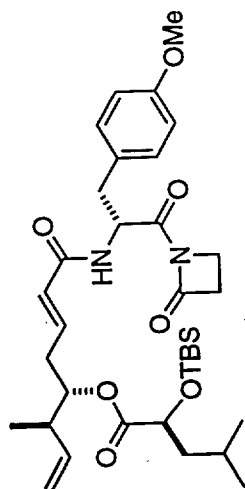
¹H NMR (400 MHz, CDCl₃)

Current Data Parameters
 NAME MJE-III-138
 EXPNO 1
 PROCNO 1

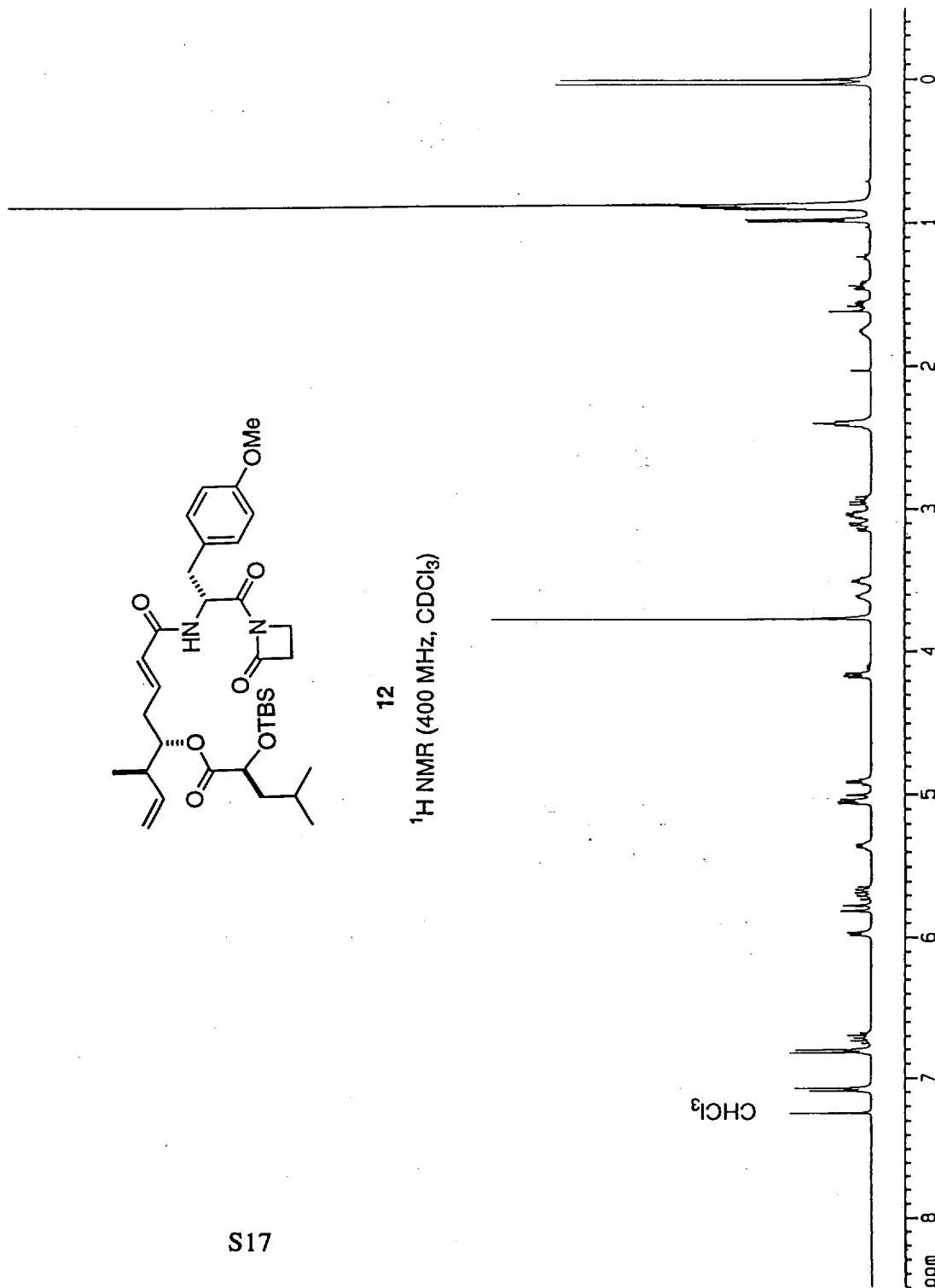
F2 - Acquisition Parameters
 Date_ 990115
 Time 17:22
 INSTRUM dr400
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 181
 DK 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.0000000 sec
 P1 7.70 usec
 DE 4.50 usec
 SF01 400.1320007 MHz
 NUC1 ¹H
 PL1 -5.00 dB

F2 - Processing parameters
 SI 16384
 SF 400.1300144 MHz
 NDX EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 8.500 ppm
 F1 3401.10 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCH 0.45000 ppm/cm
 HZCM 180.05850 Hz/cm



12

¹H NMR (400 MHz, CDCl₃)CHCl₃

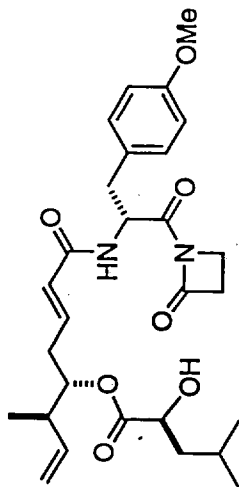
S17

Current Data Parameters
NAME sn-d-103f
EXPNO 1
PROCNO 1

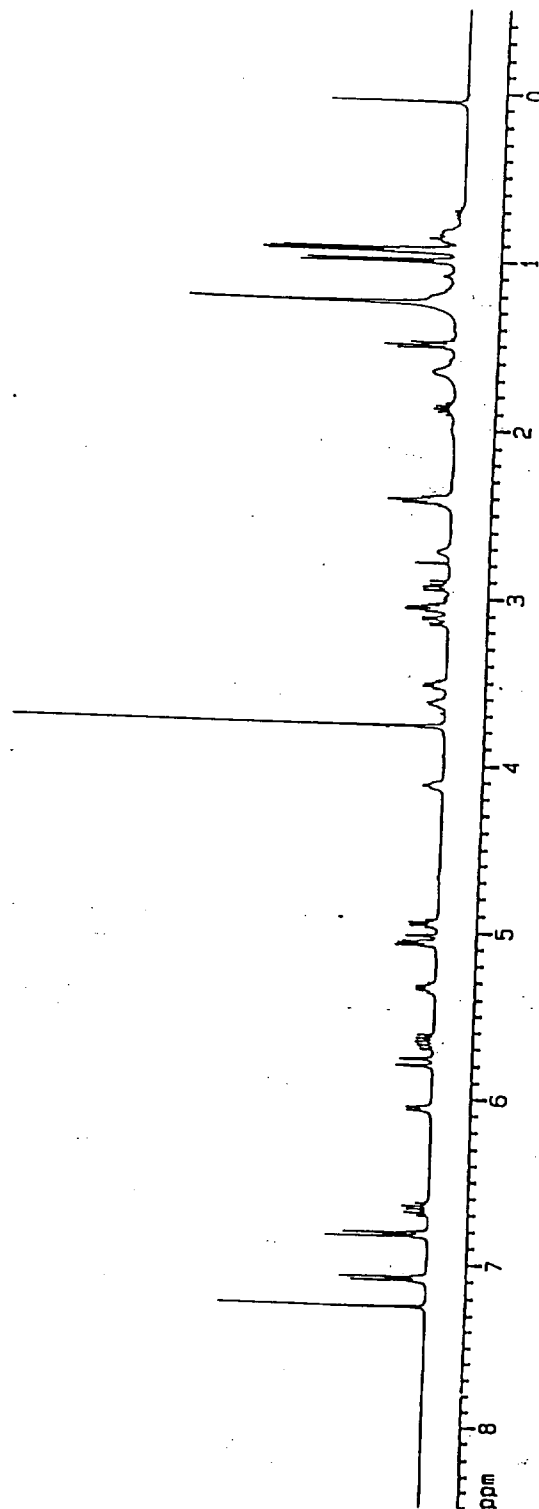
F2 - Acquisition Parameters
Date_ 990823
Time 17.37
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zg30
TO 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 256
DH 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.0000000 sec
P1 7.70 usec
DE 4.50 usec
SFO1 400.1320007 MHz
NUC1 1H
PL1 -6.00 dB

F2 - Processing parameters
SI 16384
SF 400.1300141 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
F1P 8.500 ppm
F1 3401.10 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PP4CM 0.45000 ppm/cm
HZCM 180.05850 Hz/cm



5

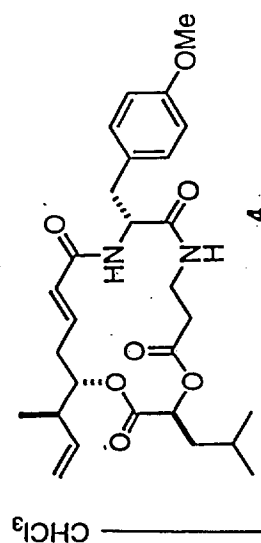
¹H NMR (400 MHz, CDCl₃)

Current Data Parameters
 NAME MJJ-III-pmac
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 1000229
 Time 17:05
 INSTRUM drx400
 PROBHD 5 mm Mxltinu
 PULPROG zg30
 TO 32768
 SOLVENT CDCl₃
 NS 71
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 256
 DH 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 P1 7.70 usec
 DE 4.50 usec
 SF01 400 1320007 MHz
 NUC1 ¹H
 PL1 -6.00 dB

F2 - Processing parameters
 SI 16384
 SF 400 1300135 MHz
 H0H EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

10 NMR plot parameters
 CX 20.00 cm
 F1P 8.500 ppm
 F1 3401.10 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCH 0.45000 ppm/c
 HZCH 180.05850 Hz/c



4
¹H NMR (400 MHz, CDCl₃)

S19

